

Invasion/migration assay

Materials

Cells from 12 well or 6 well (1x10^6 cells are enough for 1x invasion/migration)

24well plate (BD companion plate)

BD BioCoat Growth Factor Reduced MATRIGEL Invasion Chamber

BD control insert (without matrigel)

Accutase

Quick differential staining kit

Media condition

MCF10A

Assay media A (serum free)

Assay media A (serum free)	Amount	Final	
DMEM/F12 media	100ml		
BSA (35%)	286ul	0.1%	

			NII.	
LA	BOF	RAT	OF	RY

DMEM/F12 media	100ml	LABORATORY	
BSA (35%)	286ul	0.1%	
Horse serum	5ml	5%	

EGF 20ul 20ng/ml

Other cell lines growing in normal DMEM

Media A

Serum free (-FBS)

Media B

10% FBS

Procedure

A. Preparation of membrane (Only for inserts with matrigel)

- 1. Remove the foil package and put at RT until it gets warmed (Don't back to -20oC!)
- 2. Rehydrate membranes by media A (500ul for a insert and 500ul for a well)

(Not to touch the down side of membranes)

3. Incubate at 37oC for 2h (for pH equilibrium)

B. Starting assay (30min to 2h)

- 1. Wash cells with PBS
- 2. Detach cells by 500ul of accutase, and add 500ul of media A

- 3. Count cells, spin down, and re-suspend as
- 4. 200ul of cell suspension and 1.8ml of media A to adjust to 10x10^4/ml (5x10^4/500ul)
- 5. 750ul of media B in the bottom chamber
- 6. Remove media from rehydrated inserts and set them into the wells of BD 24 well plate as well as control inserts
- 7. 500ul of cell mixture in the top chamber
- 8. Incubate at 37oC 22~24h

C. Fixation and staining by Differential Quik Stain Kit

- 1. Remove media and wash inserts with DW twice
- 2. Solution A for 1min followed by DW wash twice
- 3. Solution B for 1min followed by DW wash twice
- 4. Solution C for 1min followed by DW wash twice
- 5. Remove cells on upside by scrubbing with cotton bubs
- 6. DW wash twice
- 7. Dry up inserts
- 8. Observe by microscope and count, if necessary, cut membranes and put on slide glasses